

**Identification and
Characterisation of the
Enzymes Involved in the
Biosynthetic Pathway of
Tartaric Acid in *Vitis
vinifera***

By

Crista Ann Burbidge

B. Science (Hons)

Flinders University

School of Biological Sciences

Faculty of Science and Engineering

July, 2011

Table of Contents

Summary	5
Declaration	7
Acknowledgments	8
Abbreviations	9
Chapter 1: Introduction	11
1.1 General Introduction.....	12
1.2 Tartaric Acid and Higher Plants	14
1.3 Grapevine Development, Solute Localisation and Transport	17
1.4 Tartaric Acid Biosynthetic Pathway's in Higher Plants.....	20
1.4.1 The Asc-Inclusive C4/C5 Cleavage Pathway of Vitaceae	20
1.4.2 The Asc-Inclusive C2/C3 Cleavage Pathway of Geraniaceae	24
1.4.3 The Asc-Noninclusive, Glucose Precursor Pathway of Leguminosae....	27
1.5 Enzymes of the Biosynthetic Tartaric Acid Pathway's in Vitaceae.....	31
1.5.1 Step (1a) L-Ascorbic Acid → 2-Keto-L-Gulonic Acid.....	33
1.5.2 Step (1b) D-Glucose → D-Gluconic Acid	33
1.5.3 Step (2a) 2-Keto-L-Gulonic Acid → L-Idonic Acid.....	33
1.5.4 Step (2b) D-Gluconic Acid → 5-Keto-D-Gluconic Acid	34
1.5.5 Step (3) L-Idonic Acid → 5-Keto-D-Gluconic Acid	34
1.5.6 Step (4) 5-Keto-D-Gluconic Acid → Tartaric Acid Semialdehyde	35
1.5.7 Step (5) Tartaric Acid Semialdehyde → L-Tartaric Acid.....	36
1.6 Tartaric Acid Enzymes of Bacterial Origin.....	36
1.7 Dissimilation of Tartaric Acid.....	45
1.8 Environmental Factors	50
1.9 Conclusions and Aims.....	52
Chapter 2: Materials and Methods	54
2.1 Chemicals	55
2.2 Plant Material	55
2.3 Sampling.....	55
2.4 Seasonal Data Analysis	56
2.4.1 °Brix and Fresh Weight.....	56
2.4.2 Organic Acid extraction and High-Performance Liquid Chromatography Analysis	56
2.4.2.1 Organic Acid Extraction.....	56
2.4.2.2 High Performance Liquid Chromatography.....	57
2.5 RNA Extraction.....	58
2.6 cDNA Synthesis via Reverse Transcription.....	59
2.7 Gene Expression Analysis via Quantitative Real Time Polymerase Chain Reaction.....	59
2.8 Genomic DNA Extraction from Young Leaf Tissue.....	61
2.9 Enzyme Extraction from Berry Tissue	61

2.10 Bioinformatics	62
2.11 Polymerase Chain Reaction.....	63
2.11.1 Nucleic Acid Sequencing	66
2.12 Construction of Candidate Cassette for Protein Expression	66
2.12.1 Restriction Digest of Template.....	66
2.12.2 Ligation of Candidate Open Reading Frame into Vector.....	66
2.12.2.1 pDRIVE Vector.....	66
2.12.2.2 pET-14b Vector.....	67
2.12.3 Transformation of Vector + Cassette into <i>E. coli</i>	67
2.13 Competent Cells for DNA Cloning and Protein Expression.....	67
2.14 Culture Media for Cell Growth and Protein Expression	68
2.15 Expression, Extraction and Purification of Recombinant Protein.....	69
2.15.1 Extraction of Soluble Recombinant Protein	69
2.15.1.1 Standard Extraction	69
2.15.1.2 Cell Lysis and Extraction of Soluble Proteins using BugBuster™	70
2.15.2 Protein Purification via Immobilized Metal Affinity Chromatography	70
2.15.3 Confirmation of Protein Purity via Sodium Dodecyl Sulphate	
Polyacrylamide Gel Electrophoresis	71
2.16 Kinetic Parameters of Enzyme Activity.....	71
2.17 Calculation of Enzyme Activity.....	72
2.18 Liquid Chromatography Mass Spectrometry	72

Chapter 3: Identification and Bioinformatic Analysis of Candidate Genes Encoding Enzymes Involved in Tartaric Acid Biosynthesis 73

3.1 Introduction	74
3.2 Results	76
3.2.1 Identification and Bioinformatic Analysis of Candidate Genes Involved in Tartaric Acid Biosynthesis.....	76
3.3 Discussion	92

Chapter 4: Cloning, Heterologous Expression and Analysis of Candidate Tartaric Acid Biosynthetic Enzymes 97

4.1 Introduction	98
4.2 Results	100
4.2.1 Cloning of Putative <i>Vitis vinifera</i> Tartaric Acid Biosynthetic Genes into an Expression Vector	100
4.2.2 Heterologous Expression and Purification of Recombinant <i>Vitis vinifera</i> Proteins in <i>Escherichia coli</i>	101
4.2.3 Determination of the Kinetic Parameters of Enzyme Activity.....	109
4.2.4 Optimal pH of Enzyme Activity	110
4.2.5 Favourable pH of TC59682 Catalytic Activity	111
4.2.6 Activity of Recombinant TC59682 Protein Against 2-keto-L-gulonate	113
4.2.7 Coenzyme Affinity of TC59682.....	115
4.2.8 Optimal Temperature of TC59682 Activity.....	119
4.2.9 Substrate Range of Putative Recombinantly Expressed <i>Vitis vinifera</i> Enzymes	120

4.2.10 Identification of Product Obtained from Candidate TC59682's Primary Reaction.....	124
4.3 Discussion	128
Chapter 5: Characterisation of Various Physiological and Molecular Parameters of Tartaric Acid Biosynthesis in <i>Vitis vinifera</i> cv Shiraz over the 2007/2008 Season	139
5.1 Introduction	140
5.2 Results	141
5.2.1 Fresh Weight and °Brix.....	141
5.2.2 Organic Acids Accumulation	143
5.2.3 Gene Expression Analysis utilising Qualitative Real Time PCR.....	144
5.2.4 Enzyme Extractions.....	150
5.3 Discussion	155
Chapter 6: Using Complementation Strategies to Characterise Candidates Genes Encoding Putative Members of Tartaric Acid Biosynthetic Pathway	164
6.1 Introduction	165
6.2 Results	170
6.2.1 Complementation Studies using Mutant <i>Escherichia coli</i> Strains Growing on Various Carbon Sources.....	170
6.2.2 Identification of Putative Tartaric Acid Biosynthetic Enzymes in Members of the Vitaceae Family.	175
6.2.3 Gene Expression Analysis utilising Qualitative Real Time PCR in RNAi Thompson Seedless Lines	180
6.3 Discussion	185
Chapter 7: Discussion and Future Work	190
7.1 Introduction	191
7.2 Future Research.....	192
7.3 Conclusion.....	195
Appendix	197
A.1 Analysis of the RNAi L-IdnDH knockdown Thompson Seedless Lines	198
Bibliography	203

Summary

Vitis vinifera cv Shiraz is a member of Vitaceae, one of three higher plant families in which tartaric acid (TA) accumulates to significant levels. The accumulation of TA in *V. vinifera* berry tissue is crucial for commercial wine making, preventing discolouration and spoilage due to bacterial contamination. In *V. vinifera*, TA is biosynthesised via two pathway's, the primary-ascorbate and secondary-glucose precursor pathway's. Little is known regarding the enzymes responsible for the biosynthesis of TA with L-idonate dehydrogenase (L-idonate-5-dehydrogenase) the only enzyme identified in the primary-ascorbate precursor pathway. The results presented in this study describe a bioinformatic approach to the identification of putative candidates for a 2-keto-D-gluconate reductase (possessing 2-keto-L-gulonate reductase activity) and gluconate 5-dehydrogenase suggested as being responsible for the catalysis of the second steps of the primary-ascorbate and secondary-glucose precursor pathway's respectively. *In vitro* biochemical characterisation of recombinantly expressed proteins in conjunction with *in vivo* molecular analysis was performed to support the candidates inclusion in the respective pathway's.

Comparative analysis of the *V. vinifera* genome with enzymes identified as catalysing identical reactions in bacteria identified three candidate 2-keto-D-gluconate reductases, *TC61548*, *TC59682* and *TC55752* and three candidate gluconate 5-dehydrogenases, *TC52437*, *TC58004* and *TC55097* (Gene Indices: Grape database).

All candidate genes were identified in immature berry cDNA except *TC52437*, therefore *TC52437* was not pursued further. No active recombinant protein was obtained for candidates *TC61548*, *TC58004* and *TC55097*. Kinetic analyses were performed on purified samples of the recombinant *TC59682* and *TC55752* protein. Optimal activity of recombinant *TC59682* was observed at 35°C, pH 7.5. Activity studies indicated the primary substrate of *TC59682* to be 2-keto-L-gulonate (2KGA) with a K_m of 4.67mM. The K_m for NADH was also determined as 0.77mM. A two-step assay utilising the highly-specific L-idonate dehydrogenase indicates L-idonate is a product of this reaction. The reversibility of *TC59682* was confirmed against L-

idonate in the presence of NAD^+ , at a rate 37-fold lower than the forward direction under identical conditions. Activity was also observed against ascorbate, the pathway precursor, 8.9-fold lower than that observed against 2KGA. Activity increased against all tested substrates in the presence of coenzyme NADP(H) as compared to NAD(H). Candidate TC55752 showed activity against 2KGA and ascorbate, 7.13-fold lower and 1.82 fold higher than TC59682 in the presence of coenzyme NADPH respectively.

QRT-PCR analysis of the candidate genes expression was conducted in developing *V. vinifera* cv Shiraz berries over the 2007-2008 season. The expression pattern of TC59682 strongly coincided with the biosynthesis of TA over development. TC55752's expression pattern does not indicate involvement in TA biosynthesis. Expression patterns of TC61548, TC58004 and TC55097 suggest these candidates are not involved in TA biosynthesis

Activity of extracted soluble enzymes from a subset of the sampled Shiraz berries showed an increase in activity against 2KGA over development suggesting, as recently shown with L-idonate dehydrogenase, the presence of the enzyme late in berry development.

The results presented in this study suggest TC59682 has a role in the primary TA biosynthetic pathway. Based upon primary substrate activity TC59682 will be annotated as a 2-keto-L-gulonate reductase.

Declaration

I certify that this thesis does not incorporate without acknowledgment any material previously submitted for a degree or diploma in any university; and that to the best of my knowledge and belief it does not contain any material previously published or written by another person except where due reference is made in the text.

Crista Burbidge

Acknowledgments

I wish to acknowledge my supervisors and co-supervisors Associate Professor Kathleen Soole, Dr Christopher Ford and Dr Matthew Hayes for their encouragement, advice and guidance throughout my PhD and for their critical review of this manuscript. I thank Kathleen for giving me the opportunity to undertake this research, Christopher for welcoming me into your lab and Matthew for your constant interest in my research.

To fellow PhD students Crystal Sweetman, Vivek Vijayraghavan, Vanessa Melino, Kerry Dungey, Cyd Yonker and other members of the Flinders and Waite campuses, thank you for all the invaluable advice you have given me both in the laboratory, tea room and over a cold beverage. For all their support and help keeping me focused or distracted, whichever the occasion called for. A special thank you to Crystal, whose advice and support got me through the most frustrating times of my research.

I wish to acknowledge my Mother Karen, Father Paul and Brother Neil for their constant reassurance, love and support in making this journey possible. For keeping my head above the water and always answering the phone, I would not be me without you.

Abbreviations

2KGA	2-keto-L-gulonate
2KGR	2-ketogluconate reductase
5KGA	5-Keto-D-Gluconic Acid
AGRF	Australian Genome Research Facility
AKR	aldo-keto reductase
ANK	Ankyrin
Asc	Ascorbic acid
ATP	Adenosine triphosphate
BLAST	Basic Local Alignment Search Tools
BME	β -mercaptoethanol
BSA	Bovine Serum Albumin
CAU	Corrected Area Units
cDNA	complementary DNA
DEDTC	diethyldithiocarbamate trihydrate
DHA	Dehydroascorbate
DMSO	dimethyl sulfoxide
DNA	deoxyribonucleic acid
DTT	Dithiothreitol
EDTA	ethylenediaminetetraacetic acid
EST	Expressed Sequence Tags
FAM	Fluorescein
FLC	full length clone
FW	fresh weight
G5Dh	gluconate-5 dehydrogenase
GA	D-gluconic acid
GC-MS	Gas chromatography–mass spectrometry
gDNA	Genomic DNA
GFP	Green Fluorescent Protein
HEPES	4-(2-Hydroxyethyl)piperazine-1-ethanesulfonic acid
HPLC	High Performance Liquid Chromatography
IA	L-idonic acid
IMAC	immobilized metal affinity chromatography
IPTG	Isopropylthiogalactoside
KCN	potassium cyanide
kDa	kilodalton
KEGG	Kyoto Encyclopedia of Genes and Genomes
LB	Luria-Bertani
LC-MS	Liquid chromatography–mass spectrometry
L-IdnDH	L-idonate dehydrogenase
LN	liquid nitrogen
MA	Malic acid
MDH	Malate dehydrogenase
MEB	Modified Elution Buffer
mETC	mitochondrial electron transport chain

mRNA	Messenger RNA
MS	mass spectrometry
MSB	Modified Sonication Buffer
MWB	Modified Wash Buffer
MWL	Molecular weight ladder
NAD ⁺	Nicotinamide adenine dinucleotide
NADH	Nicotinamide adenine dinucleotide, reduced
NADP ⁺	Nicotinamide adenine dinucleotide phosphate
NADPH	Nicotinamide adenine dinucleotide phosphate, reduced
NCBI	National Centre for Biotechnology Information
NoC	no coenzyme
NoE	no enzyme
NoS	no substrate
nt	Nucleotide
OA	Organic Acid
OG	octyl gallate
ORF	open reading frame
OxA	oxalic acid
PCR	Polymerase Chain Reactions
PEB	protein extraction buffer substituted for purified enzyme
PEG	polyethylene glycol
PI	isoelectric point
PMSF	phenylmethanesulfonyl fluoride
PVDF	Polyvinylidene Difluoride
PVP	Polyvinylpyrrolidone
PVPP	Poly(vinylpyrrolidone)
QRT-PCR	Quantitative Real Time Polymerase Chain Reaction
RE	Restriction enzymes
RNA	Ribonucleic acid
RNAi	RNA interference
SARDI	South Australian Research and Development Institute
SDS-PAGE	Sodium Dodecyl Sulphate Polyacrylamide Gel Electrophoresis
TA	Tartaric acid
TB	Terrific Broth
TC	tentative consensus sequence
TCEP	<i>tris</i> (2-carboxyethyl)phosphine
TEMED	N,N,N',N'-Tetramethylethylenediamine
ThA	L-threonic acid
TrB	Transformation Buffer
TSAD	tartaric semialdehyde dehydrogenase
Ubq	Ubiquitin
WT	wild type