Dynamics of phytoplankton in relation to tuna fish farms in Boston Bay and near-shore Spencer Gulf, South Australia

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Declaration

I certify that this thesis does not incorporate without acknowledgement any material previously submitted for a degree or diploma in my university. To the best of my knowledge and belief it does not contain any material previously published or written by another person except where due reference is made in the text.

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Rosemary Paxinos December 10, 2007

Abstract

Interest in the effect of fish farming practices on the marine environment has arisen because there is concern that the wastes that fish farms produce may be contributing to eutrophication in coastal areas and the problem of harmful algal blooms. The focus of this thesis is an examination of phytoplankton distribution and abundance in relation to tuna fish farms in Boston Bay and near-shore Spencer Gulf. This is the first study in South Australia to define the short-term biomass fluctuations of chlorophyll and in vivo fluorescence, identify phytoplankton species distribution and abundance, including two potentially toxic dinoflagellates, and describe patch distribution relative to tuna fish farms in Boston Bay and the near-shore waters of Spencer Gulf. An ecological interpretation of phytoplankton distribution and abundance is determined and shows that community composition was different in lower Spencer Gulf compared to Boston Bay and upper Spencer Gulf sites. Pico- and nanophytoplankton were often the most abundant organisms. Diatoms and gymnoids were most common. Season and currents predominantly influenced the distribution of phytoplankton in Boston Bay and Spencer Gulf. Individual species may be influenced by inputs from the fish farms.

Chlorophyll levels were different between the Spencer Gulf and Boston Bay sites and no differences were recorded, using mean levels of chlorophyll, between tuna cages and controls. Chlorophyll levels were higher east of Boston Island in autumn of 1999. Chlorophyll levels appeared to show a slight increase between years. This may have been an anomalous natural variation and future research may investigate this in the long term. In addition, Principal Components Analysis (PCA) was used to investigate differences between treatments and the functional grouping model supported an ecological interpretation of the factors from the PCA. A total of 131 taxa of phytoplankton were identified in this study. The 14 dominant taxa were used in the PCA and of these, 9 were diatoms. Phytoplankton abundance was not different between tuna cages and controls. However, when examining individual species, Karenia mikimotoi was more prevalent at tuna cages, close to shore, east and west of Boston Island than at other sites. PCA showed how different species bloomed together and were seasonal. Karenia brevis and K. mikimotoi featured predominantly in the PCA with K. brevis the dominant organism during summer and autumn along with Gyrodinium spp. and smaller gymnoids. K. brevis blooms were most likely influenced by water temperatures and fixation of nitrogen from a Trichodesmium erythaeum bloom. K. mikimotoi bloomed bimodally and may be influenced by ammonia excreted from fish from the tuna farms but, on the other hand, may be limited by the high salinities of South Australian waters. Currents in the region distribute both organisms.

The final aspect of this study assessed finer temporal and spatial sampling using directional transects around tuna cages and controls using *in-vivo* fluorescence and size fractionated chlorophyll. The chlorophyll *a* sampling showed little spatial variability within a site in the 1000 m² that the sampling area covers but far greater temporal variability (days). In contrast, fluorescence `mapping' expands the window of variability both

iv

Abstract

spatially (within a site) and temporally (along transects and between days). This has given a spatial definition, which is unavailable from a single point sample, and thereby leaves room for much greater interpretation. Small patches are evident from the fluorescence mapping where this is impossible to detect from the single point samples. Therefore, the fluorescence `mapping' and patch definition show that the trend is widespread (spatially) and quite persistent (temporally) around the fish farm area.

Size fractionated chlorophyll samples provided further insight into phytoplankton dynamics in this study where diatoms were favored over dinoflagellates and were responsible for the larger fraction of chlorophyll found at the tuna cage one (TC1) site. We suggest that seasonal fluctuations, high nutrient input from the farm activities and turbulence may be responsible for the different chlorophyll/fluorescent structures found at TC1. Future research may look at the long-term regional impact on phytoplankton size structure, biomass and communities from fish farm activities.

As a good part of this journey involved counting phytoplankton using the Utërmohl technique, a short paper, published in the *Journal of Plankton Research,* on reducing the settling time of this method, is presented in Appendix 2.

V

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Contents

Declaration	ii
Abstract	iii
Acknowledgements	vi
Figures	xii
Tables	XV

Chapter 1

Introduction	1
Fish farming in South Australia	2
Studies around tuna farms in South Australia	6
Fish farming and waste produced	9
Contribution of oceanography and hydrography to phytoplankton	
and fish farm ecology	13
Spatial and temporal distribution of phytoplankton around fish farms	15
Broad aims	23

Chapter 2

Introduction	24
Functional classification of species and habitat	26
Aims	27
Methods	27
Results	32
Water quality	32
Dissolved oxygen	34
Ammonia	35
Chlorophyll a	35
Statistical analysis	36
Phytoplankton abundance and distribution	39
Principal components analysis	39
Discussion	53
Chapter 3	

Introduction	63

Karenia brevis	63
Karenia mikimotoi	65
Aims	67
Methods	67
Results	70
Description of Karenia brevis 'look-a-likes'	70
Description of Karenia mikimotoi 'look-a-likes'	71
General distribution	71
Discussion	83
Chapter 4	
	90
Continuous flow <i>in vivo</i> fluorescence	92
Size fractionated chlorophyll a	93
Aims	94
Methods	95
Experimental design	95
In vivo fluorescence, chlorophyll a size fractionation	96
and abundance	
Data analysis	99
Results	102
Environmental conditions	102
Total chlorophyll a	102
Chlorophyll $a - 5 \mu m$ fraction	107
Chlorophyll $a - 0.45 \mu m$ fraction	108
Fluorescence signal along directional transects	109
Statistical analysis of fluorescence transect data	110
Response of <i>in vivo</i> fluorescence within 50 m of the tuna	113
cages	
Phytoplankton abundance	114
Spectral analysis	114
Patch definition	116
Discussion	118

Chapter 5	
Summary of chapters	127
Chapter 2	127
Chapter 3	129
Chapter 4	130
Discussion	132
Literature cited	149
Appendix 1	167
Appendix 2	172

Figures

Chapter 1	
Figure 1: Map showing Boston Bay, Port Lincoln, South Australia.	3
Chapter 2	
Figure 1: Map of Boston Bay (BB) and Spencer Gulf (SG), South	
Australia, showing all sampling sites including tuna cages (TC)	
and controls (C).	29
Figure 2: Mean (<u>+</u> 1SD) of (a) temperature, (b) salinity, (c) pH, (d)	
DO, and (e) ammonia at tuna cages and controls, 1997-99.	33–34
Figure 3: Mean chlorophyll a LOG10 (<u>+</u> 1SE) calculated across all	
sites and each sampling time, for treatments and controls.	36
Figure 4: Mean (\pm 1SE) of factor scores for PC1 and date for tuna	
cages (TC) and controls (C) in Boston Bay and Spencer Gulf.	
	41
Figure 5: Mean (\pm 1SE) of factor 2 and date, for tuna cages (TC)	
and controls (C) in Boston Bay and Spencer Gulf.	42
Figure 6: Mean (\pm 1SE) of factor scores for PC3 and date, for	
tuna cages (TC) and controls (C) in Boston Bay and Spencer	43
Gulf.	
Figure 7: Mean (\pm 1SE) of factor scores for PC4 and date, for	
tuna cages (TC) and controls (C) in Boston Bay and Spencer	43
Gulf.	
Figure 8: Mean algal cells/L of taxa positively associated with	44
PC1.	
Figure 9: Mean cells/L of taxa negatively associated with PC1.	45
Figure 10: Mean algal cells/L of taxa positively associated with	46
PC2.	
Figure 11: Mean algal cells/L of taxa positively and negatively	
associated with PC3.	47
Figure 12: Mean algal cells/L of taxa positively and negatively	40
associated with PC4.	48

Chapter 3

Figure 1: Map of Boston Bay (BB) and Spencer Gulf (SG), South	
Australia, showing all sampling sites including tuna cages (TC)	
and controls (C).	69
Figure 2: <i>Karenia brevis</i> 'look-alike'.	71
Figure 3: <i>Karenia mikimotoi</i> 'look-alike'.	72
Figure 4: Mean K. brevis (Kb) and K. mikimotoi (Km) cells/L over	
all sites and all sampling months in 1997/98.	73
Figure 5: Mean <i>K. brevis</i> (Kb) and <i>K. mikimotoi</i> (Km) cells/L (<u>+</u>	
1SE) by area.	75
Figure 6: Average temperatures (°C) and mean cells/L for <i>K</i> .	
brevis (Kb) and K. mikimotoi (Km) over all sites and sampling	76
months.	
Figure 7: Mean <i>K. brevi</i> s (Kb) cells/L by areas.	77
Figure 8: Mean <i>K brevis</i> (Kb) cells/L (<u>+</u> 1SE) for tuna cages (TC)	
and controls (C) over all sites and sampling months.	78
Figure 9: <i>K. mikimotoi</i> (Km) mean cells/L (<u>+</u> 1SE) by areas.	79
Figure 10: Mean <i>K. mikimotoi</i> (Km) cells/L (<u>+</u> 1SE) for tuna cages	
(TC) and controls (C) over all sites and sampling months.	80
Figure 11: Mean ammonia (μ g/L) levels at tuna cages (TC) and	
controls (C) compared to mean K. brevis (Kb) and K. mikimotoi	
(Km) cells/L for all sites and sampling months.	81
Figure 12: Mean temperatures (°C) compared to mean <i>K. brevis</i>	
(Kb) and K. mikimotoi (Km) cells/L for all sites, over all sampling	82
months.	

Chapter 4

Figure 1: Map of Boston Bay, South Australia showing tuna cage	
1 (TC1) and 2 (TC2), and control sites 1 (C1) and 2 (C2).	96
Figure 2: Diagrammatic representation of the direction of the tuna	
cage and control transects.	97
Figure 3: Calibration curve for the conversion of fluorescence	
(relative units) to chlorophyll a, derived by chlorophyll a extraction	
of water samples taken from the outlet of the flow-through	

fluorometer.	98
Figure 4: Mean Chl a samples taken at tuna cages (TC) and	
controls (C) on day 1 at 0, 50, and 100 m (<u>+</u> 1 SE).	104
Figure 5: Mean Chl a samples taken at tuna cages (TC) and	
controls (C) on day 2 at 0, 50, and 100 m (<u>+</u> 1 SE).	105
Figure 6: Mean Chl a samples taken at tuna cages (TC) and	
controls (C) on day 3 at 0, 50, and 100 m (<u>+</u> 1 SE).	105
Figure 7: Mean ChI a samples taken at tuna cages (TC) and	
controls (C) on day 4 at 0, 50, and 100 m (<u>+</u> 1 SE).	106
Figure 8: Mean Chl a samples taken at tuna cages (TC) and	
controls (C) on day 5 at 0, 50, and 100 m (<u>+</u> 1 SE).	106
Figure 9: Mean ChI <i>a</i> 5μ m fraction for each sampling day at tuna	
cages (TC) and controls (C) (<u>+</u> 1 SE).	107
Figure 10: Mean ChI <i>a</i> 0.45 μ m fraction for each sampling day at	
tuna cages (TC) and controls (C) (<u>+</u> 1 SE).	108
Figure 11: Example of a time plot for in vivo fluorescence	
(RU=relative units) transects and temperature ($^{\circ}$ C) for TC2 on day	
3, in the westerly direction (TC=tuna cage; W1 and W2=westerly	
replicate transects).	110
Figure 12: Example of power spectra (f is frequency) of the	
fluorescence time series (log/ RU=relative units) and temperature	
time series for north 1 transect at C2 (control) on day 5, shown	
with their best-fitting lines in a log-log plot.	115
Figure 13: Example of patch definition at TC1 (a) and C1 (b) on	
day 1 pooled and detrended fluorescent (RU=relative units)	
transect data.	116

Tables

Chapter 2

Table 1: Taxa of phytoplankton with abbreviation and the principal	
component number.	39
Table 2: Loadings of each included taxon for the four principal	
components extracted and percent variation explained.	40
Table 3: Mean cell counts (cells/L) for all phytoplankton groups	
(when picoplankton are excluded) displayed as a percentage by	
area.	51

Chapter 4

Table 1: Mean (<u>+</u> 1SE) of total Chl <i>a</i> (pooled 0, 50 and 500 m) for all	
tuna cage sites over 5 sampling days.	103
Table 2: Multiple comparisons of total Chl <i>a</i> (pooled 0, 50 and 500	
m) for all tuna cage sites over 5 sampling days (n=45).	104
Table 3: Example of range, mean $(\pm 1SE)$ and sample size of	
fluorescence (RU=relative units) for all sites on day 1, north one (N1)	
transect only (TC=tuna cage; C=control).	112
Table 4: Results of univariate analysis of variance comparing	
between effects of the first 50 m of the fluorescence transects and	
between effects of last 50 m at tuna cages (TC) and controls (C).	