

Characterisation of X Chromosomal Short Tandem Repeat Markers for Forensic Use

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Declaration Page

I certify that this thesis does not incorporate without acknowledgment any material previously submitted for a degree or diploma in any university; and that to the best of my knowledge and belief it does not contain any material previously published or written by another person except where due reference is made in the text.

The opinions or assertions presented hereafter are the private views of the author and should not be construed as official or as reflecting the views of the Department of Defense, its branches, the US Army Medical Research and Materiel Command or the Armed Forces Medical Examiner System.

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Toni Marie Diegoli *1/15/14*
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Abstract

The use of X chromosomal short tandem repeat (STR) markers has been greatly increasing in the forensic setting. The marker system offers the potential to provide information in addition to that which is obtained from autosomal STR systems currently employed at forensic and paternity laboratories and in the courtroom. In certain scenarios, markers on the X chromosome may be the only means of obtaining the required information. Any investigated relationship situation where at least one female is involved will benefit from the use of X STRs, which can be applied to cases of missing persons, criminal incest, immigration, deficiency paternity or other questioned relationships. In-depth characterisation of the marker system is the first step in maximizing the power of this additional tool in the forensic arsenal.

Using guidelines set forth within the 1991 report of the International Society for Forensic Genetics (ISFG) relating to the use of DNA polymorphisms, all aspects of the feasibility of routine X STR use were evaluated. Two mini-X chromosomal STR multiplexes capable of amplifying 15 total markers (DXS6795, DXS9902, DXS8378, DXS7132, DXS6803, DXS6789, DXS7424, DXS101, GATA172D05, DXS7130, GATA165B12, HPRTB, GATA31E08, DXS10147, and DXS7423) were developed and optimised for use in the investigation and characterisation of allele nomenclature, allele/genotype frequencies, mutation rates, and linkage between markers. To simplify the transition into routine use, the assays utilised techniques and instrumentation already present in most forensic laboratories as well as analyses familiar to forensic scientists. Several large sample sets from four U.S. populations (African Americans, U.S. Asians, U.S. Caucasians, and U.S. Hispanics) were studied to address the lack of relevant data available for the United States, where the use of X STRs remains limited. Much of the knowledge gained, however, is universally applicable.

Though commercial kits are available that probe a wide variety of genetic markers on both the Y chromosome and the autosomes, there is only one commercial kit currently manufactured assaying markers on the X chromosome, the Investigator Argus X-12 kit (QIAGEN, Hilden, Germany). Due to patent and intellectual property issues between the United States and European STR kit manufacturers, the

kit cannot be sold or marketed in the U.S. at this time. Nonetheless, four markers overlapping with those present in the Argus X-12 kit were included in the optimised multiplexes and were the subject of an extensive concordance study. With greater than 99% concordance across 975 alleles, comparability of the data was established for these four markers while the utility of the kit with U.S. populations was evaluated for the first time.

This extensive developmental validation study investigated each aspect of the X STR marker system that would require consideration before implementation by a forensic laboratory. The optimised assays were found to be robust and the markers discriminating, while the mutation rate was estimated with high accuracy and the extent of linkage between the 15 markers was thoroughly evaluated. The combination of the results obtained as part of this study form the foundation upon which the introduction and routine use of X STRs may be built.

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Abbreviations

μL	Microliter
μM	Micromolar
A	Adenine
AFDIL	Armed Forces DNA Identification Laboratory
BLAST	Basic Local Alignment Search Tool
BLAT	BLAST-like Alignment Tool
bp	Base pair(s)
BSA	Bovine serum albumin
C	Cytosine
CEPH	Centre d'Etudes du Polymorphisme Humaine
CI	Confidence interval
cM	Centimorgan
cm	Centimeter(s)
CODIS	Combined DNA Index System
Ct	Cycle threshold
DAB	DNA Advisory Board
DF	Dilution factor
dGTP	Deoxyguanosine triphosphate
dH ₂ O	Distilled water
DNA	Deoxyribonucleic acid
dNTP	Deoxyribonucleotide triphosphate
EDNAP	European DNA Profiling Group
EDTA	Ethylenediaminetetraacetic acid
EMPOP	EDNAP mtDNA Population Database
FamID	Family identification number
FID	Father's identification number
F _{ST}	Fixation index
G	Guanine
H(exp)	Expected heterozygosity
H(obs)	Observed heterozygosity
INDEL	Insertion/deletion marker
IPC	Internal PCR control
IRBO	Institutional Review Board Office
ISFG	International Society for Forensic Genetics
ISFH	International Society for Forensic Hameogenetics
kb	Kilobase(s)
kV	Kilovolts
LD	Linkage disequilibrium
LG	Linkage group
LISA	Laboratory Information Systems Applications
LOD	Logarithm of the odds
LR	Likelihood ratio
Mb	Megabase(s)
MEC	Mean exclusion chance
MECI	Mean exclusion chance in trios involving daughters
MECII	Mean exclusion chance in father/daughter duos
MgCl ₂	Magnesium chloride
MID	Mother's identification number
MIXplex	Mixture multiplex

mM	Millimolar
MRMC	U.S. Army Medical Research and Materiel Command
mtDNA	Mitochondrial DNA
NCBI	National Center for Biotechnology Information
ng	Nanograms
NIGMS	National Institute of General Medical Sciences
NIST	National Institute of Standards and Technology
°C	Degrees Celcius
p (HWE)	P value of the exact test for Hardy-Weinberg equilibrium
PCR	Polymerase chain reaction
PDf	Power of discrimination in females
PDm	Power of discrimination in males
pg	Picograms
PIC	Polymorphism information content
PID	Patient identification number
Ref.	Reference
RF or Θ	Recombination fraction (or recombination rate)
SNP	Single nucleotide polymorphism
SRY	Sex-determining region of the Y chromosome
STR	Short tandem repeat
STRBase	Short Tandem Repeat Database
SWGDM	Scientific Working Group on DNA Analysis Methods
T	Thymine
TLE	Tris-low-EDTA buffer
T_m	Melting temperature
U.S.	United States
UCSC	University of California Santa Cruz
YHRD	Y Chromosome Haplotype Reference Database
Z	LOD score

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Appendix C. Additional Publications

- C1. Toni M. Diegoli, Michael D. Coble, Development and characterization of two mini-X chromosomal short tandem repeat multiplexes, *Forensic Sci. Int. Genet.* 5 (2011) 415-421.
- C2. Toni M. Diegoli, Lejla Kovacevic, Naris Pojskic, Michael D. Coble, Damir Marjanovic, Population study of fourteen X chromosomal short tandem repeat loci in a population from Bosnia and Herzegovina, *Forensic Sci. Int. Genet.* 5 (2011) 350-1.
- C3. Toni M. Diegoli, Chapter 5: Forensic X Chromosomal Short Tandem Repeat Typing, in: Moses Schanfield and Dragan Primorac, (Eds.), *Forensic DNA Applications: An Interdisciplinary Perspective*, Taylor & Francis group, *prepared winter 2012 and submitted February 2013 for publication late 2013; not included here due to size constraints.*
- C4. Toni M. Diegoli, Adrian Linacre, Michael D. Coble, Characterization of X chromosomal short tandem repeat markers for forensic use, *Forensic Sci. Int. Gene. Suppl.* (2013), <http://dx.doi.org/10.1016/j.fsigss.2013.10.074>.
- C5. Toni M. Diegoli, Adrian Linacre, Michael D. Coble, A gonosomal marker multiplex to aid in mixture interpretation, *Forensic Sci. Int. Gene. Suppl.* (2013), <http://dx.doi.org/10.1016/j.fsigss.2013.10.095>.

References