HERITABLE INFLUENCES IN OXYGEN-INDUCED RETINOPATHY

Peter van Wijngaarden MBBS (Hons)



Thesis submitted for the degree of Doctor of Philosophy January 2006

Faculty of Health Sciences School of Medicine Flinders University of South Australia Adelaide, Australia For Amber

TABLE OF CONTENTS

SUMMARY OF THESIS	IX
PUBLICATIONS ARISING FROM THIS THESIS	XI
DECLARATION	XII
ACKNOWLEDGEMENTS	XIII
ABBREVIATIONS	XV
1. CHAPTER 1: INTRODUCTION	1
1.1. OVERVIEW	2
1.2. RETINAL ANATOMY	2
1.2.a. The retinal pigment epithelium	4
1.2.b. The sensory retina	5
1.2.c. The retinal vasculature	6
1.2.c.1 Retinal vascular anatomy	7
1.2.c.2 Retinal vascular physiology – retinal oxygenation	9
1.3. VASCULAR DEVELOPMENT	11
1.3.a. Blood vessel structure	11
1.3.b. Vasculogenesis and angiogenesis	12
1.3.c. Mechanisms of angiogenesis	13
1.4. OCULAR VASCULARIZATION IN DEVELOPMENT	21
1.4.a. Development of the choroidal and hyaloid circulations	21
1.4.b. Overview of retinal vascular development	22
1.4.c. Mechanisms of retinal vascular development	24
1.4.c.1 Historical insights	24
1.4.c.2 Current concepts of retinal vascular development: vasculogenesis	
1.4.c.3 Current concepts of retinal vascular development: angiogenesis	
1.5. RETINOPATHY OF PREMATURITY	
<i>1.5.a. Clinical features and the classification of retinopathy of prematurity</i>	
1.5.b. ROP epidemiology and prognostic factors	
1.5.c. Treatment of ROP	
1.5.d. The role of oxygen in the pathogenesis of ROP	40
1.5.d.1 Oxygen – cause or cure?	
1.5.d.2 Oxygen induces retinopathy in animal models of ROP	
1.5.a. Genetic risk factors for neovescularization and POP	
1.5.c. Generic risk juciors for neovascularization and KOT	
1.6 HVPOTHESIS AND AIMS	

2.	СНАР	TER 2: MATERIALS & METHODS	53
2	2.1. N	IATERIALS	54
	2.1.a.	Water	54
	2.1.b.	General chemicals	54
	2.1.c.	Enzymes	54
	2.1.d.	PCR primers	54
	2.1.e.	Experimental animals	58
	2.1.6	Ethical considerations	
	2.1.6	2.2 Rats	
	2.1.f.	Miscellaneous materials	60
2	E.2. E	SUFFERS AND SOLUTIONS	61
	2.2.a.	Buffered formalin	61
	2.2.b.	Chrome alum-subbed microscope slides	61
	2.2.c.	DEPC-H ₂ O	61
	2.2.d.	DEPC-NaCl	61
	2.2.e.	Electrophoresis gel DNA/RNA loading buffer	61
	2.2.f.	Eosin	61
	2.2.g.	Ethidium bromide agarose plates	61
	2.2.h.	Haematoxylin solution	62
	2.2.i.	Ink perfusate	62
	2.2.j.	PBS (10X)	62
	2.2.k.	Sodium hydroxide stock 10 M	62
	2.2.l.	TBE buffer (10x)	62
2	2.3. N	IOLECULAR TECHNIQUES	63
	2.3.a.	DNA quantification	63
	2.3.b.	Agarose gel electrophoresis	63
	2.3.c.	RNA extraction	63
	2.3.d.	DNAseI treatment of RNA extracts	64
	2.3.e.	RNA quantification and qualitative assessment	65
	2.3.f.	cDNA synthesis	65
	2.3.g.	Preparation of the standard cDNA sample	66
	2.3.h.	Preparation of test cDNA samples	67
	2.3.i.	Polymerase chain reaction primer design	68
	2.3.j.	General precautions for polymerase chain reaction	
	2.3.k.	Endpoint reverse transcription-polymerase chain reaction	70
	2.3.l.	Quantitative real-time reverse transcription-polymerase chain reaction	71
	2.3.m.	Agarose gel PCR-product purification	73
	2.3.n.	PCR product sequencing	73

	2.4. A	NIMAL AND TISSUE TECHNIQUES	74
	2.4.a.	Conventional histology	74
	2.4.b.	Ink perfusion of rats	74
	2.4.b	.1 Perfusion device	74
	2.4.b	.2 Preparing rats for perfusion	
	2.4.b	.3 Priming the perfusion device	75
	2.4.b	.4 Perfusion technique	77
	2.4.c.	Oxygen-induced retinopathy	79
	2.4.c	.1 Oxygen chamber	
	2.4.c	.2 Oxygen exposure protocol	
	2.4.c	.3 Retinal dissection and flat-mounting	
	2.4.c.	.4 Isolectin histochemistry	
	2.4.c.	.5 Image analysis of labelled retinae	
	2.4.d.	Mechanical ventilation of neonatal rats	89
	2.5. S'	TATISTICAL ANALYSIS	92
	2.5.a.	Retinal avascular area and vascular morphology	
	2.5.b.	Real-time PCR data	
3.	СНАР	FER 3: RESULTS – STRAIN COMPARISON OF OXYGEN-INDUCED	
	RETI	NOPATHY	93
	21 D	NTRODUCTION	04
	21a	Quantiau	
	5.1.u.	Diele farsteren fan OID	
	<i>3.1.0</i> .	Risk jactors for OIR	
	3.2. K	ESULIS	
	3.2.a.	Strain comparisons of retinal vascularization in normoxia	
	<i>3.2.b</i> .	Strain comparisons of retinal vascularization following exposure to cyclic h	yperoxia104
	3.2.b	.1 Avascular retinal area	
	3.2.b	.2 Vascular morphology	
	3.2.c.	Body mass	
	<i>3.2.d</i> .	Respiratory function	
	3.2.e.	A hereditary basis for susceptibility to OIR: cross-breeding studies	114
	3.2.e.	.1 The susceptibility of the offspring of F344 x DA crosses to OIR	
	3.2.e.	.2 The susceptibility of the back-cross offspring to OIR	
	3.3. D	ISCUSSION	119
4.	CHAP	FER 4: RESULTS – RETINAL GENE EXPRESSION	129
	4.1. II	NTRODUCTION	130
	4.1.a.	Vascular endothelial growth factor	130
	4.1.b.	Vascular endothelial growth factor receptor-2	131
	4.1.c.	Angiopoietin 2 & Tie2	
	41d	Cvclooxvgenase-2	134
	···	-,,8	

4.1.e.	Erythropoietin	134
4.1.f.	Insulin-like growth factor-1	135
4.1.g.	Pigment epithelium-derived factor	136
4.2. 0	OVERVIEW OF THE EXPERIMENTAL DESIGN	139
4.2.a.	Methodological validation	140
4.2.a	a.1 Confirmation of primer specificity	140
4.2.a	a.2 Determination of primer amplification efficiency	148
4.2.a	a.3 Optimisation of experimental conditions	150
<i>4.2.b</i> .	Relative quantification of gene expression	
4.2.c.	Normalisation of gene expression	151
4.2.0	c.1 Reference gene selection and validation	151
4.2.0	Normalised retinal gene expression	161
4.2.d.	Intra-run and inter-run variation	
4.2.e.	Experimental design and methodological validation: concluding remarks	164
4.3. I	NTER-ANIMAL VARIATION IN GENE EXPRESSION	166
4.4. S	STRAIN DIFFERENCES IN RETINAL GENE EXPRESSION FOLLOWING EXPO)SURE
]	ГО CYCLIC HYPEROXIA AND RELATIVE HYPOXIA	171
4.4.a.	VEGF mRNA expression	
4.4.b.	VEGFR-2 mRNA expression	173
4.4.c.	PEDF mRNA expression	173
4.4.d.	Ang2 mRNA expression	
4.4.e.	Tie2 mRNA expression	
4.4.f.	EPO mRNA expression	
4.4.g.	IGF-1 mRNA expression	
4.4.h.	COX-2 mRNA expression	179
4.4.i.	Interpretation of the observed changes in gene expression – correlation with strain	1
	differences in retinal vascular area	
4.4.i	.1 Strain differences in gene expression — exposure day 14	
4.4.i	.2 Strain differences in gene expression — exposure day 18	
4.5. S	STRAIN DIFFERENCES IN RETINAL GENE EXPRESSION DURING CYCLIC	
I	HYPEROXIA	
4.5.a.	Experimental design	
4.5.b.	VEGF mRNA expression during cyclic hyperoxia	
4.5.c.	VEGFR-2 mRNA expression during cyclic hyperoxia	
4.5.d.	PEDF mRNA expression during cyclic hyperoxia	
4.5.e.	Ang2 mRNA expression during cyclic hyperoxia	
4.5.f.	EPO mRNA expression during cyclic hyperoxia	
4.5.g.	Summary of strain differences in angiogenic factor gene expression during the cou	urse of
-	cyclic hyperoxia	

4.6.	RETINAL GENE EXPRESSION IN THE BACKCROSS PROGENY OF DA AN	D F344
	RAT STRAINS FOLLOWING CYCLIC HYPEROXIA	197
4.7.	DISCUSSION	200
4.7.a	Possible mechanisms for the observed strain differences in retinal gene expression	ion between
	F344 and DA rats during cyclic hyperoxia	
4.	7.a.1 VEGF	
4.	7.a.2 VEGFR-2	
4.	7.a.3 Ang2	
<i>4.7.b</i>	b. Differences in retinal gene expression provide a basis for the strain differences	in retinal
	vascularization in rat OIR	214
4.7.c	Reconciling the lack of a strain difference in PEDF expression during cyclic hyperbolic hyperbolic strain difference in PEDF expression during cyclic strain during cyclic str	peroxia
	with the difference evident after cyclic hyperoxia	
4.7.a	I. The failure of retinal vascularization despite angiogenic factor expression	216
4.7.e	The implications of inter-animal variation in gene expression on the differences	observed
	between pooled samples	217
4.7.f.	Closing remarks	218
5. CHA	APTER 5: DISCUSSION	219
5.1.	INTRODUCTION	220
5.2.	STUDY FINDINGS	220
5.2.a	Strain comparisons	
5.2.b	The heritability of susceptibility to oxygen-induced retinopathy	221
5.2.c	Quantitative retinal gene expression	
5.2.d	l. Differential vascularization of the central and peripheral retina	
5.2.e	Study limitations	
5.2.f.	Integration of study findings	
5.3.	AN OVERVIEW OF RELATED STUDIES	
5.3.a	A comparison of the susceptibility of Brown Norway and Sprague Dawley rats t	to OIR228
5.3.b	Rat strain differences in retinal vascular permeability in OIR and in diabetes	230
5.3.c	Lessons from the anterior segment: genetic factors may regulate sensitivity to V	EGF and
	<i>bFGF</i>	
5.3.d	l. Strain differences in murine OIR	
5.3.e	Parallels in tumour angiogenesis	
5.3.f.	The broader implications of rat strain-related heterogeneity in the susceptibility	, to OIR 240
5.3.g	Bronchopulmonary dysplasia and ROP: more than meets the eye?	241
5.4.	FUTURE DIRECTIONS	244
5.5.	FINAL COMMENTS	245
APPENDI	IX 1: INBRED RAT STRAINS	247
A1.	INBRED RAT STRAINS	

APPEND	IX 2: PCR PRODUCT SEQUENCE DATA	249
A2.	SEQUENCE DATA	
A2	1. ARBP	
A2.2	2. Ang2	
A2	3. COX-2	
A2.4	4. EPO	253
A2	5. HPRT	253
A2.0	5 PEDF	254
A2.1	7. RNAP2	255
A2.8	8. Tie2	255
A2.	9. VEGF	256
A2	10. VEGFR-2	
A2.11.	IGF-1 sequence	257
	IGF-1A	258
	IGF-1B	258
APPEND	IX 3: REAL-TIME RT-PCR MATHEMATICS	260
A3.	MATHEMATICAL CONSIDERATIONS IN REAL-TIME RT-PCR	
<i>A3</i> .	I. PCR amplification kinetics	
A3	<i>I.a</i> Using PCR kinetics for relative quantification: the delta-delta Ct method	
A3	l.b. The delta Ct method of quantification	
A3.2	2. PCR efficiency calculation	
APPEND	IX 4: NORMALISATION OF GENE EXPRESSION DATA	265
A4.1.	BACKGROUND	
A4.2.	SELECTION OF CANDIDATE REFERENCE GENES	
A4.3.	REFERENCE GENE EXPRESSION STABILITY AND NORMALISATION USING MULTIPLE R	EFERENCE
	GENES	
A4.3.A	. GENORM INTERNAL GENE-STABILITY MEASUREMENT	271
BIBLIO	GRAPHY	272

SUMMARY OF THESIS

Retinopathy of prematurity, a disease characterised by aberrant retinal vascular development in premature neonates, is a leading cause of blindness and visual impairment in childhood. This work sought to examine differences in the susceptibility of inbred rat strains to oxygen-induced retinopathy, a model of human retinopathy of prematurity. The overriding aim was to identify genetic factors in rats that might be generalisable to humans.

Newborn rats of six different strains were exposed to alternating cycles of hyperoxia and relative hypoxia for fourteen days. Rats were removed to room air and killed for analysis immediately, to assess oxygen-induced retinal vascular attenuation, or four days later to evaluate the extent of hypoxia-induced vasoproliferation. Whole flat-mounted retinae were stained with fluorophore conjugated isolectin GS-IB4, and measurement of vascular area was conducted using fluorescence microscopy and video-image analysis. A hierarchy of susceptibility to the inhibitory effects of cyclic hyperoxia and relative hypoxia on postnatal retinal vascularization was identified for the rat strains studied. Susceptibility to vascular attenuation was predictive of the subsequent risk of vascular morphological abnormalities. Cross-breeding experiments between susceptible and resistant strains demonstrated that the susceptible phenotype was dominantly inherited in an autosomal fashion. These studies confirmed an association between ocular pigmentation and retinopathy risk, however the finding of differential susceptibility amongst albino rat strains implicated factors in addition to those associated with ocular pigmentation.

Quantitative real-time reverse transcription-polymerase chain reaction was used to compare the retinal expression of angiogenic factor genes in susceptible and resistant strains with the aim of identifying a genetic basis for the strain difference. Eight angiogenic factor genes were selected for study: vascular endothelial growth factor (VEGF); VEGF receptor 2; angiopoietin 2; Tie2; pigment epithelium-derived factor; erythropoietin; cyclooxygenase-2 and insulin-like growth factor-1. The most notable difference between strains was the expression of vascular endothelial growth factor

(VEGF) during the cyclic hyperoxia exposure period – higher VEGF expression was associated with relative resistance to retinopathy. Other differences in retinal angiogenic factor gene expression between strains, such as higher expression of VEGF receptor 2 and angiopoietin 2 in resistant strains, appeared to be secondary to those in VEGF. Following cyclic hyperoxia, the expression pattern of angiogenic factor genes changed – messenger RNA levels of hypoxia-induced genes, including VEGF, VEGF receptor 2, angiopoietin 2 and erythropoietin, were significantly higher in those strains with larger avascular areas, than in those strains that were relatively resistant to retinopathy. These findings provide firm evidence for hereditary risk factors for oxygen-induced retinopathy in the rat. Differences in the regulatory effects of oxygen on VEGF expression appear to be central to the risk of retinopathy. The potential relevance of these hereditary factors is discussed in the context of the human disease.

PUBLICATIONS ARISING FROM THIS THESIS

- van Wijngaarden, P., Coster, D.J. and Williams, K.A, *Inhibitors of Ocular Neovascularization: Promises and Potential Problems*. Journal of the American Medical Association, March 23/30, 2005. 293(12): p. 1509-1513.
- van Wijngaarden, P., Coster, D.J., Brereton, H.M., Gibbins, I.L. and Williams, K.A, *Strain-Dependent Differences in Oxygen-Induced Retinopathy in the Inbred Rat.* Investigative Ophthalmology and Visual Science, April, 2005. 46(4): p. 1445-1452.

DECLARATION

I certify that this thesis does not incorporate without acknowledgement any material previously submitted for a degree or diploma in any university; and that to the best of my knowledge and belief it does not contain any material previously published or written by another person except where due reference is made in the text.

Signed:

Date:

Peter van Wijngaarden

ACKNOWLEDGEMENTS

In performing this work I have drawn extensively on the expertise and assistance of others. I express my sincere gratitude to all of the individuals who have aided me in my endeavours.

Specifically, I thank:

- Kirsty Marshall for preparing ocular sections for conventional histology and for her assistance with animal handling.
- Ray Yates, Theresa Fischer, Stuart Lisk and the staff of the animal housing facility for their assistance with animal handling and with animal experimentation.
- Anne-Louise Smith and staff of the Biomedical Engineering Department for constructing the oxygen chamber and for tending to its on-going maintenance.
- George Mayne, Damian Hussey, Michael Michael, Shiwani Sharma, Pam Sykes and Lesley Snell for their assistance with quantitative real-time reverse transcription polymerase chain reaction.
- Dan Peet (University Adelaide, Department of Biochemistry) and Greg Goodall (Institute of Medical and Veterinary Science, Adelaide) for their advice regarding the interpretation of the transcriptome changes identified by quantitative polymerase chain reaction.
- Oliver van Wageningen for sequencing polymerase chain reaction products.
- Ian Gibbins and Kylie Lange for their assistance with statistical analyses. Ian Gibbins also provided helpful advice regarding fluorescence microscopy and image analysis.
- Andrew Bersten, Hilde de Smet and Malgosia Krupa for their assistance with the respiratory experiments.
- Bren Gannon for advice regarding the construction of an ink-perfusion device
- Michelle Lewis for advice about retinal histology.

I am particularly indebted to my research supervisors – Keryn Williams, Helen Brereton, Doug Coster and Ian Gibbins – for their direction, their support and their friendship. Their passion for scientific research and for methodological rigour is contagious; and their tireless dedication to teaching and mentoring is admirable. I have been blessed to have received such remarkable support!

In a similar vein, I thank all of the members of the Ophthalmology Department at Flinders University for their friendship and assistance. Joyce Moore, Lyn Harding, Kirsty Marshall, Claire Jessup, Paul Badenoch, David Dimasi, Melinda Tea, Marian Turner, Matt Wenham, Alix Farrall, Scott Standfield, Margaret Philpott, Doug Parker, Claude Kauffman and Shiwani Sharma have all been bastions of support and will remain dear friends. I am also grateful for the friendship of John Oliver, Stuart Perry and John Woodall.

My four-year solo sojourn in Adelaide came at a great cost to my family. I will remain forever grateful for the sacrifices made by Marijcke and Pieter, Eric and Cath, Nicky and Frazer, Bill and Marcia, Kyla and Simon, Ward, and especially Amber, in allowing me to pursue my dream. My parents instilled in me the self-belief to embark on this work and equipped me with the drive to persevere in the face of difficulty – for this I am thankful. To my wife Amber, I owe the greatest debt of gratitude – her unconditional support and encouragement have been central to my successes. As a small gesture of my appreciation I dedicate this work to Amber.

ABBREVIATIONS

\leq	less than or equal to
≥	more than or equal to
~	approximately
#	number
°C	degrees Celsius
μg	microgram (10^{-6} g)
μl	microlitre (10 ⁻⁶ l)
μΜ	micromolar (10 ⁻⁶ M)
μm	micrometer (10 ⁻⁶ m)
А	adenine
aa	amino acid
a/bFGF	acidic/basic fibroblast growth factor
Ang 1	angiopoietin 1
Ang 2	angiopoietin 2
AP-1/-2	activator protein-1/-2
ARBP	Acidic Ribosomal Phosphoprotein
ARNT	aryl hydrocarbon receptor nuclear translocator (HIF-1 β)
ARVO	Association for Research in Vision and Ophthalmology
ATP	adenosine triphosphate
BM	Bruch's membrane
bp	base pairs
С	cytosine
cDNA	complementary DNA
cGMP	cyclic-guanosine monophosphate
cm	centimetre
COX 2	cyclooxygenase 2
CRYO-ROP	the Multicentre Trial of Cryotherapy for Retinopathy of Prematurity
Da	Dalton
DA	Dark Agouti rat strain
DAG	diacylglycerol

DAO2	Dark Agouti rats exposed to cyclic hyperoxia and relative hypoxia for the first two days of life – day two follows a 24 hour period of relative hypoxia
DAO3	Dark Agouti rats exposed to cyclic hyperoxia and relative hypoxia for the first three days of life – day three follows a 24 hour period of hyperoxia
DAO8	Dark Agouti rats exposed to cyclic hyperoxia and relative hypoxia for the first 8 days of life – day 8 follows a 24 hour period of relative hypoxia
DAO9	Dark Agouti rats exposed to cyclic hyperoxia and relative hypoxia for the first 9 days of life – day 9 follows a 24 hour period of hyperoxia
DAO14	Dark Agouti rats exposed to cyclic hyperoxia and relative hypoxia for the first 14 days of life
DAO18	Dark Agouti rats exposed to cyclic hyperoxia and relative hypoxia for the first 14 days of life, followed by four days of sustained relative hypoxia in room air
DARA14	Dark Agouti rats exposed to room air for the first 14 days of life
ddH ₂ 0	double distilled water
DEPC	diethylpyrocarbonate
DNA	deoxyribonucleic acid
dNTP	dinucleotide triphosphate
DTT	dithiothreitol
ECM	extracellular matrix
EDTA	ethylene-diamine-tetraacetic-acid
EGF	epidermal growth factor
ELM	external limiting membrane
eNOS	endothelial nitric oxide synthetase
EPO	erythropoietin
ETDRS	Early Treatment of Diabetic Retinopathy Study
EtOH	ethanol
ETROP	the Early Treatment for Retinopathy of Prematurity Randomized Trial
F _(x,y)	F statistic (degrees of freedom, error)
F344	Fischer 344 rat strain
F344O2	Fischer 344 rats exposed to cyclic hyperoxia and relative hypoxia for the first two days of life – day two follows a 24 hour period of relative hypoxia

F344O3	Fischer 344 rats exposed to cyclic hyperoxia and relative hypoxia for the first three days of life – day three follows a 24 hour period of hyperoxia
F344O8	Fischer 344 rats exposed to cyclic hyperoxia and relative hypoxia for the first 8 days of life – day 8 follows a 24 hour period of relative hypoxia
F344O9	Fischer 344 rats exposed to cyclic hyperoxia and relative hypoxia for the first 9 days of life – day 9 follows a 24 hour period of hyperoxia
F344O14	Fischer 344 rats exposed to cyclic hyperoxia and relative hypoxia for the first 14 days of life
F344O18	Fischer 344 rats exposed to cyclic hyperoxia and relative hypoxia for the first 14 days of life, followed by four days of sustained relative hypoxia in room air
F344RA14	Fischer 344 rats exposed to room air for the first 14 days of life
FAK	focal adhesion kinase
FasL	Fas-ligand (CD95L)
FGF	fibroblast growth factor
Fig	figure
flk-1	foetal-liver kinase-1 (VEGFR-2)
flt-1	fms-like tyrosine kinase-1 (VEGFR-1)
g	gram
g	gravity
G	guanine
#G	# gauge
GAPDH	glyceraldehyde 3-phosphate dehydrogenase
GCL	ganglion cell layer
GH	growth hormone
GOI	gene of interest
GS-IB4	Griffonia simplicifolia type I isolectin B4-Alexa 488™ conjugate
HPRT	hypoxanthine guanine phophoribosyl transferase
HIF-1/-2	hypoxia inducible factor-1/-2
hr	hour
HRE	hypoxia response element
HuR	Hu protein R

HW	Hooded Wistar rat strain
ICAM-1	intercellular adhesion molecule-1
Ig	immunoglobulin
IGF-1	insulin-like growth factor-1
IL	interleukin
ILM	inner limiting membrane
IM	intramuscular
INL	inner nuclear layer
IP	intraperitoneal
IPL	inner plexiform layer
iU	international units
Kb	kilobases
kDa	kilo Daltons (10 ³ Da)
KDR	kinase domain receptor (VEGFR-2)
Kg	kilogram (10 ³ gram)
1	litre
LEW	Lewis rat strain
М	molar
m	metre
MAPK	mitogen-activated protein kinase
mg	milligram (10 ⁻³ g)
MHC	major histocompatibility complex
min	minutes
MIP-2	macrophage inhibitory peptide-2 (IL-8)
ml	millilitre (10 ⁻³ l)
mm	millimetre(10^{-3} m)
mM	millimolar (10 ⁻³ M)
mmHg	millimetres mercury
MMP	matrix metalloproteinase
mRNA	messenger ribonucleic acid
MW	molecular weight
n	number/sample size

NaCl	sodium chloride
NADPH	nicotinamide-adenine dinucleotide phosphate
NFL	nerve fibre layer
ng	nanogram (10 ⁻⁹ g)
NH&MRC	National Health and Medical Research Council of Australia
No.	number
NRP-1	neuropilin-1
NSW	New South Wales
NTC	no template control
OIR	oxygen-induced retinopathy
ONL	outer nuclear layer
OPL	outer plexiform layer
ORP150	oxygen-regulated protein-150
P#	postnatal day #
PAF	platelet activating factor
PBS	Dulbecco's A physiologic balanced salt solution
PCO ₂	partial pressure of carbon dioxide
PCR	polymerase chain reaction
PDGF	platelet derived growth factor
PECAM-1	platelet-endothelial cell adhesion molecule-1
PEDF	pigment epithelium derived factor
pg	picogram (10 ⁻¹² gram)
pI	isoelectric point
PI3-kinase	phosphatidylinositol 3-kinase
PlGF	placental growth factor
pmol	picomoles (10 ⁻¹² moles)
PO ₂	partial pressure of oxygen; oxygen tension
RNA	ribonucleic acid
RNAP2	RNA polymerase 2
ROP	retinopathy of prematurity
RPE	retinal pigment epithelium
rpm	revolutions per minute

rRNA	ribosomal RNA
RT	room temperature
RT1	rat MHC Class II
RT-	reverse transcriptase-free; negative control cDNA
RT-PCR	reverse transcription-polymerase chain reaction
sec	second
SA	South Australia
SD	standard deviation
sFlt	soluble fms-like tyrosine kinase
SNP	single nucleotide polymorphism
SPD	Sprague Dawley rat strain (conventionally abbreviated SD. SPD in this thesis to avoid confusion with the abbreviation for standard deviation)
SPDO14	Sprague Dawley rats exposed to cyclic hyperoxia and relative hypoxia for the first 14 days of life
SPDO18	Sprague Dawley rats exposed to cyclic hyperoxia and relative hypoxia for the first 14 days of followed by four days of sustained relative hypoxia in room air
SPDRA14	Sprague Dawley rats exposed to room air for the first 14 days of life
Т	thymine
T _A	annealing temperature
TBE	tris borate EDTA
TGF α / β	transforming growth factor- α /- β
Tie2	tyrosine kinase with Ig and epidermal growth factor homology domain receptor 2: receptor for angiopoietin-1 & -2
T _m	melting temperature
TNF α	tumour necrosis factor-α
TUNEL	terminal deoxynucleotidyl transferase (TdT)-mediated deoxyuridine triphosphate (dUTP)-biotin nick end labelling
U	units
USA	United States of America
UV	ultraviolet light
V	volt
V	version; volume
\mathbf{v}/\mathbf{v}	unit volume per unit volume

VA	visual acuity
VCAM 1	vascular cell adhesion molecule 1
VEGF	vascular endothelial growth factor (VEGF A unless otherwise specified)
VEGF _x	VEGF, isoform _x (_x denotes number of amino acid residues)
VEGFR-1	vascular endothelial growth factor receptor-1
VEGFR-2	vascular endothelial growth factor receptor-2
VEGFR-3	vascular endothelial growth factor receptor-3
VHL	von Hippel-Lindau protein
VIC	Victoria
VPF	vascular permeability factor
V _T	tidal volume
w/v	unit weight per unit volume
WA	West Australia
WF	Wistar-Furth rat strain
WG	weeks of gestation
Х	times / multiplication factor