Preclinical *in vitro* and *in vivo* effects of purified and synthetic bioactive compounds from marine mollusc *Dicathais orbita* on colorectal cancer: Cancer prevention and toxicity study



By

Babak Esmaeelian, DVM

Submitted in fulfillment of the requirements for the degree of Doctor of Philosophy.

School of Biological Sciences, Faculty of Science and Engineering, Flinders

University, Adelaide, South Australia.

July, 2013

Table of Contents

Table of Figures vi

Data Tables ix

Abstract x

Declaration xv

Prefacexvi

Acknowledgments xviii

Abbreviations xx

1. General introduction 1

1.1 Colorectal cancer 2

1.2 Biology of colorectal cancer 3

1.3 Treatment and prognosis of colorectal cancer 7

1.4 Prevention of colorectal cancer 11

1.4.1 Screening tests: 11

1.4.2 Impact of life style in colorectal cancer prevention 13

1.5 Natural products benefits and their anticancer properties 15

1.5.1 Marine natural products and molluscs as a potential source of anticancer compounds 17

1.5.2 Muricidae molluscs as a source of bioactive compounds 20

1.6 Preclinical studies for anticancer drug development 23

1.6.1 Colon cancer cell lines 24

1.6.2 Cell viability assays 24

1.6.3 Apoptosis or programmed cell death versus necrosis 25

1.6.4. Flow cytometry for detection of apoptosis and cell cycle analysis 28

1.6.5. In vivo models for colon cancer prevention 29

1.6.6 Toxicity testing 30

1.7 Aims 32

1.8 Thesis structure 33

2. Optimizing the purification of tyrindoleninone and 6-bromoisatin and testing the proliferative effect of Vitamin E stabilized tryindoleninone on HT29 cells 36

2.1 Introduction 37
2.2 Material and methods 39
2.2.1 Extraction and separation of compounds by thin-layer chromatography (TLC)39
2.2.2 Flash chromatography and chemical analysis 40
2.2.3 Antioxidant assay 41
2.2.4 Cell viability assay42
2.3 Results and discussion 43
2.3.1 TLC separation 43
2.3.2 LC/MS analysis 46
2.3.3 Antioxidant effects on extract stability 49
2.3.4 Synergistic effects of Vitamin E on cell viability 52
2.4 Conclusion 55
3. Purified brominated indole derivatives from Dicathais orbita induce apoptosis and cell cycle arrest in colorectal cancer cell lines57
3.1 Abstract 58
3.2 Introduction 59
3.3 Material and methods 62
3.3.1 Egg mass extraction, purification62
3.3.2 Chemical analysis63
3.3.3 Cell culture64
3.3.4 MTT viability assay and cell morphology 64
3.3.5 Combined caspase 3/7, membrane integrity and cell viability assays 65
3.3.6 Flow cytometric detection of apoptosis 66
3.3.7 Cell cycle analysis66
3.3.8 Statistical analysis67
3.4 Results and discussion 67
3.4.1 Chemical analysis and bioassay guided fractionation 67
3.4.2 Biological activity of the D. orbita compounds 74
3.5 Conclusion 86
3.6 Acknowledgments86

4. Brominated indoles from a marine mollusc extract prevent early stage colon 87 92 4.3.1 Hypobranchial gland extraction, flash column chromatography and chemical 93 4.3.3 Measurement of apoptosis using hematoxylin staining 94 4.3.4 Determination of epithelial proliferation (immunohistochemical staining) 4.3.5 Histopathological evaluation of the liver 96 4.3.6 Blood analysis (hematology and biochemistry)96 97

4.4 Results 97

4.3.7 Statistical analysis

cancer formation in vivo.

4.3 Materials and Methods

4.3.2 In vivo rodent model

92

4.1 Abstract 88

4.2 Introduction

analysis

97 4.4.1 Chemical analysis

4.4.2 In vivo model - General observations 99

88

4.4.3 Apoptotic index and crypt height 101

4.4.4 Proliferation index 104

4.4.5 Histopathology evaluation of the liver 107

4.4.6 Liver enzyme alteration 109

4.4.7 Hematology and other serum biochemical alterations 110

4.5 Discussion 113

4.6 Acknowledgments121

5. 6-Bromoisatin found in muricid mollusc extracts inhibits colon cancer cell proliferation and induces apoptosis, preventing early stage tumor formation in a colorectal cancer rodent model 122

5.1 Abstract 123

5.2 Introduction 124

5.3 Material and Methods 127

5.3.1 Synthetic 6-bromoisatin and chemical analysis 127

5.3.2 In vitro experiments using HT29 colorectal cancer cells 128 95

5.3.3 In vivo model for early stage colon cancer prevention 129

5.3.4 Liver enzymes, blood biochemistry and hematology 131

5.3.5 Statistical analysis 131

5.4 Results and Discussion 131

5.4.1 Chemical analysis 131

5.4.2 In vitro apoptosis, necrosis and cell viability: 132

5.4.3 In vivo mouse model 137

5.5 Conclusions 149

5.6 Acknowledgments150

6. Final discussion and future directions 151

6.1 Summary of research 152

6.2 The use of natural antioxidants in combinatorial anticancer drug therapies 153

6.3 Cell culture assays for drug screening 154

6.3.1 Cell viability assays 156

6.3.2 Apoptosis tests 160

6.4 In vivo trials for drug screening 162

6.5 Toxicity studies for drug screening 164

6.6 Pharmaceuticals from isatins 167

6.7 Potential development of 6-bromoisatin as a combinatorial chemotherapy for colorectal cancer 169

6.8 Potential development of 6-bromoisatin as a new product for colorectal cancer prevention or treatment 171

6.9 Final conclusion 173

7. Appendix: Analysis of variance (ANOVA) and Post Hoc Tukey HSD tables 174

7.1 In vitro result analysis from Chapter 2 175

7.2 In vitro result analysis from Chapter 3 177

7.3 In vivo result analysis from Chapter 4 189

7.4 In vitro and in vivo result analysis from Chapter 5 209

References 224

Table of Figures

Figure 1.1 Adenocarcinoma progression model for colorectal cancer by Harrison
and Benziger (2011)
Figure 1.2 Dicathais orbita, the source of biologically active secondary metabolites,
adapted from Westley and Benkendorff (2008)
Figure 1.3 Three apoptosis mechanism including two main intrinsic and extrinsic
pathways with the third perforin/granzyme pathway (Elmore, 2007) 27
Figure 2.1 Thin layer chromatography of the crude extract from the hypobranchial
gland of <i>D. orbita</i> using different solvent systems from low to high polarity45
Figure 2.2 Liquid chromatography- mass spectrometry analysis of purified/semi-
purified fraction from hypobranchial gland extract of <i>D. orbita</i> 47
Figure 2.3 LCMS analysis of the semi-purified tyrindoleninone and tyrindolinone
from <i>D. orbita</i> after 24 h oxygen exposure
Figure 2.4 Cell viability of HT29 cells (a) and Caco2 cells (b) treated with different
concentrations of semi-purified fraction containing tyrindoleninone and
tyrindolinone
Figure 3.1 LC-MS analysis of extract from <i>D. orbita</i> egg capsules71
Figure 3.2 MTT viability results of <i>D. orbita</i> egg mass crude extract (CE) and all
fractions collected from flash column chromatography (Frac 1-7)72
Figure 3.3 GC-MS chromatogram of fractions from the egg masses extract of the
Australian muricid, <i>D. orbita</i> 73
Figure 3.4 Effects of <i>D. orbita</i> egg mass crude extract (CE), purified
tyrindoleninone (TYR) and semi-purified 6-bromoisatin (6-BRO) on HT29 and
Caco2 cells80

Figure 3.5 HT29 cells at 400x magnification under the Olympus inverted		
microscope		
Figure 3.6 Flow cytometric analysis of HT29 cells		
Figure 3.7 Cell cycle analysis using propidium iodide (PI) staining and flow		
cytometry		
Figure 4.1 Experimental timeline for the two week AOM- induced rodent model for		
colon cancer prevention		
Figure 4.2 Liquid chromatography- mass spectrometry analysis of a) crude extract		
from hypobranchial gland of D. orbita b) purified tyrindoleninone and c) semi-		
purified 6-bromoisatin		
Figure 4.3 Apoptotic response in the epithelial cells of crypts in the distal colon after 14 day oral gavage of different concentrations of tyrindoleninone (TYR), semi-		
purified 6-bromoisatin (6-BRO) and crude extract (CE) 102		
Figure 4.4 Apoptosis in distal colon crypts of a) AOM control and b) a mouse administered 0.1 mg/g semi-purified 6-bromoisatin		
Figure 4.5 Mean (±S.E.) crypt height in the distal colon of control mice and		
treatment groups including different concentrations of purified tyrindoleninone		
(TYR), semi-purified 6-bromoisatin (6-BRO) and crude extract (CE)104		
Figure 4.6 Proliferation index in the distal colon of control groups and animals treated with different concentrations of purified tyrindoleninone (TYR), semi-purified 6-bromoisatin (6-BRO) and crude extract (CE) for 14 days105		
Figure 4.7 Proliferative activity using an antibody specific for the ki-67 antigen in		
distal colon crypts of a) AOM control and b) mouse administered 0.1 mg/g semi-		
purified 6-bromoisatin (6-BRO) for 14 days106		
Figure 4.8 Microscopic images of sectioned liver tissue108		

Figure 4.9 Liver enzymes aspartate aminotransferase (AST), alanine
aminotransferase (ALT) and alkaline phosphatase (ALP) levels in serum109
Figure 5.1 1H NMR spectrum of synthetic 6-bromoisatin on the Bruker Avance III
400 MHz spectrometer in deuterated acetonitrile
Figure 5.2 Effects of synthetic 6-bromoisatin on HT29 cells
Figure 5.3 HT29 cells at 400x magnification under the Olympus inverted
microscope
Figure 5.4 Apoptotic response and crypt height in the distal colon of mice after 14
day oral gavage with different concentrations of 6-bromoisatin139
Figure 5.5 Apoptosis in the basal crypt cells of the distal colon of mice 6 h post
AOM injection
Figure 5.6 Proliferative activity of distal colonic epithelial cells in mice 6 h after
AOM injection
Figure 5.7 Proliferation index in the distal colon of oil control mice and mice treated
with different concentrations of 6-bromoisatin
Figure 5.8 Liver enzymes aspartate aminotransferase (AST), alanine
aminotransferase (ALT) and alkaline phosphatase (ALP) levels in serum (U/L) of
control mice and mice treated with different concentrations of 6-bromoisatin145

Data Tables

Table 1.1 Status of marine-derived natural products in anticancer clinical trials,
adapted from Petit and Biard (2013)
Table 2.1 Retention time and mass values of major compounds in the crude
chloroform extract from the hypobranchial gland (HG) and the egg mass (EM) of D.
orbita by LCMS analysis
Table 2.2 Percent composition (relative to the sum of brominated compounds) of
semi-purified tyridoleninone and tyrindolinone from <i>D. orbita</i>
Table 4.1 Comparison of mean (±S.E.) progressive body weight (g) in control and
treatment mice on different experimental days100
Table 4.2 Hematology results from mouse blood samples at the end of a two week
period of oral gavage111
Table 4.3 Biochemical results of mouse serum samples
Table 5.1 Comparison of mean (±S.E.) progressive body weight (g) in controls and
mice treated with different concentrations of 6-bromoisatin
Table 5.2 Plasma biochemistry and blood hematology from mice in the oil only
control group and mice treated with different concentrations of 6-bromoisatin147
Table 6.1 Summary of the different effects of the crude extract and compounds from
D. orbita from a suite of in vitro and in vivo experiments

Abstract

Marine indole-based compounds, including precursors and isomers of the ancient purple dye, Tyrian purple, are known for their biological activity. In particular, the precursors 6-bromoisatin and tyrindoleninone from the Australian whelk, Dicathais orbita are compounds that have gained specific interest over the past few years for their anticancer effects in several cancer cell lines. Previous *in vivo* studies in mice by administration of D. orbita extract has indicated the potential for these bioactive compounds to prevent colon cancer, but with possible idiosyncratic liver toxicity. Therefore, purification of the most likely bioactive compounds (tyrindoleninone and 6-bromoisatin) from *D. orbita* could be helpful to enhance the anticancer properties and potentially reduce the toxicity associated with the crude extract. Synthetic 6bromoisatin is commercially available, so testing the pure synthetic compound will also help confirm any activity associated with this compound. Tyrindoleninone is a compound which can be easily oxidized to other components. Therefore, stabilizing it by using antioxidants might be beneficial to increase its bioactive effects. The objective of this project was to optimize the purification of tyrindoleninone and 6bromoisatin and examine the effects of these compounds, along with crude extract from D. orbita, on colorectal cancer in vitro and in vivo. The toxicity of these compounds and extracts was also assessed in vivo to establish the safety of these compounds in the body system.

To optimize the purification of tyrindoleninone and 6-bromoisatin, initially the compounds were separated from the extract using thin layer chromatography (TLC) using a gradient of hexane, dichloromethane and methanol. Then flash

chromatography was used to purify these compounds using the most effective solvent system from TLC. The purified compounds were analysed using liquid chromatography/mass spectrometry (LC/MS) to confirm their identity and purity. The chemical composition of crude extracts from egg masses and hypobranchial glands were also compared by LC/MS and found to contain a very similar percent composition of the main brominated compounds. In order to inhibit the degradation of tyrindoleninone, a fraction containing tyrindoleninone and tyrindolenine was exposed to oxygen overnight in the presence of two antioxidants, Vitamin A and Vitamin E, and then reanalysed by LC/MS. The synergic anti-proliferative effect of tyrindoleninone with the most effective antioxidant was then tested on HT29 cells using 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl) tetrazolium bromide (MTT) assay. The antioxidant experiment showed that 0.1 % Vitamin E was the most effective antioxidant for inhibiting tyrindoleninone degradation, but it failed to increase the cytotoxic effect of tyrindoleninone on HT29 cells and in fact appeared to provide some protection against the cytotoxic properties of tyrindoleninone.

In the next *in vitro* experiments, an egg mass extract was used for purification of the bioactive compounds with the optimised flash silica chromatography method. Bioassay guided fractionation was performed to identify the compounds with the greatest antiproliferative effects against colon cancer cells. The identity of the main bioactive compounds was confirmed by LC/MS, GC/MS and NMR as tyrindoleninone (>99% purity) and 6-bromoisatin (90% purity). These compounds were then tested for cytotoxic, apoptotic or necrotic effects using MTT, caspase 3/7 and membrane integrity assays respectively, on HT29 and Caco2 cells. The apoptotic effects of the bioactive compounds were confirmed by flow cytometry using Annexin-V-FITC and

PI staining. Cell cycle analysis was also performed on HT29 cells treated with the most bioactive compound. The 3-(4,5-dimethythiazol-2-yl)-2,5-diphenyl tetrazolium bromide (MTT) assay showed that semi-purified 6-bromoisatin inhibited the viability of both cell lines (IC₅₀= 100 μ M). The fraction containing 6-bromoisatin activated caspase-3 and -7 enzymes in Caco2 and HT29 cells at approximately 100 μ M (0.025 mg/mL) and 200 μ M (0.05 mg/mL) respectively, much lower concentrations than those required to cause LDH release and necrosis (~1000 to ~2000 μ M). Flow cytometry showed that semi-purified 6-bromoisatin (~200 μ M) induced 77.6% apoptosis in HT29 cells. Cell cycle analysis showed the accumulation of 25.7% of HT29 cells treated with semi-purified 6-bromoisatin (~100 μ M) in G2/M phase. The other compound, tyrindoleninone, was also found to inhibit the proliferation of Caco2 cells (IC₅₀= 98 μ M) and HT29 (IC₅₀= 390 μ M). Caspase-3 and -7 activity significantly increased only in HT29 cells treated with 195 μ M (0.05 mg/mL) tyrindoleninone. LDH was released in both cell lines treated with high concentrations of tyrindoleninone.

In an *in vivo* trial, the effects of the purified tyrindoleninone and semi-purified 6bromoisatin, along with the crude extract were tested for prevention of colorectal cancer in a two week mouse trial to determine whether these compounds can enhance the acute apoptotic response to genotoxic carcinogens (AARGC). The antiproliferative effects of the extract and purified/semi-purified compounds were also tested by immunohistological examination using Ki-67 antibody. To evaluate any possible toxicity of the compounds, mouse general health, behavior, body weight and liver weight were assessed. Liver damage was also tested using histopathology and also biochemistry by measuring liver enzymes (ALT, AST and ALP) in the serum. Some other biochemical and also hematological blood tests were performed to evaluate any other toxicity or side effects in blood and kidney. Semi-purified 6bromoisatin (0.05 mg/g) was found to be the most bioactive compound in the crude extract capable of enhancing the apoptotic index in distal colon of mice. Tyrindoleninone did not increase the apoptotic index significantly. Semi-purified 6bromoisatin did not show any toxic effect on liver, as indicated by no significant difference in the liver enzymes in comparison to the controls. In contrast, tyrindoleninone caused an increase in AST level compared to the saline control and also caused a reduction in red blood cell counts.

In my last experiment, pure synthetic 6-bromoisatin was tested for *in vitro* anticancer activity and prevention of the colorectal cancer using the same *in vivo* model. Administration of pure synthetic 6-bromoisatin to the mice, confirmed the results from the semi-purified 6-bromoisatin, with a significant increase in apoptosis at 0.05 mg/g, without any sign of toxicity in the liver or blood cells. However, a decrease in the potassium levels in the blood indicated the possibility of a diuretic effect associated with synthetic 6-bromoisatin.

This research confirmed the anticancer effects of 6-bromoisatin against two colorectal cancer cell lines *in vitro*, as well as the potential cancer preventative effects *in vivo* based on the ability to induce apoptosis in DNA damaged cells. This supports the potential development of this molluscan extract or natural 6-bromoisatin as a nutraceutical for chemoprevention of colorectal cancer. In addition, synthetic 6-bromoisatin is a promising lead for further pharmaceutical development for prevention of this disease. However, future studies in longer term animal models are required to

confirm that the early stage prevention of tumors by apoptosis in DNA damaged cells by 6-bromoistain does prevent the formation of actual tumors at the later developmental stages.

Declaration

I certify that this thesis does not incorporate without acknowledgement any material previously submitted for a degree of diploma in any university; and to the best of my knowledge and belief it does not contain any material previously published or written by another person except where due reference is made in the text.

Babak Esmaeelian

Preface

Parts of the work presented in this thesis have been submitted for publication, or are currently in preparation for submission for publication. In this thesis, Chapters 3-5 are in manuscript format and hence there is some necessary repetition in the methods and introduction sections. Chapters 2, 3 and 5 are in the format more typical for chemistry journals (e.g. *Marine Drugs*) with combined results and discussion, whereas Chapter 4 is in the format more typical for cancer journals (e.g. *Cancer Letters*), with separate results and discussion sections. All references are listed at the end of the thesis to eliminate repetition,

Manuscripts for publication:

Chapter 3. <u>Esmaeelian, B</u>, Benkendorff, K, Johnston, MR and Abbott, CA "Purified brominated indole derivatives from *Dicathais orbita* induce apoptosis and cell cycle arrest in colorectal cancer cell lines" This chapter has been published as a research paper in Marine Drugs on 11 October 2013.

Chapter 5. <u>Esmaeelian, B</u>, Benkendorff, K, Le Leu, R and Abbott, CA "Bromoisatin found in muricid mollusc extracts inhibits colon cancer cell proliferation and induces apoptosis, preventing early stage tumor formation in a colorectal cancer rodent model" This chapter has been published as a research paper in Marine Drugs on 24 December 2014.

Chapter 2. <u>Esmaeelian, B</u>, Abbott, CA and Benkendorff, K "Optimizing the purification of tyrindoleninone and 6-bromoisatin and testing the proliferative effect of stabilized tryindoleninone on HT29 cells" A part of this chapter (The optimization of purification) is under preparation to *BMC Complementary Medicine* but the other part (Stabilization of tyrindoleninone) will go to another Journal.

Chapter 4. <u>Esmaeelian, B</u>, Benkendorff, K, Le Leu, RK and Abbott, CA "Brominated indoles from a marine mollusc extract prevent early stage colon cancer formation *in vivo*" Under preparation for submission to *Cancer Letters*

Conference presentations:

<u>Esmaeelian, B</u>, Benkendorff, K and Abbott, CA, "Preclinical testing of purified Muricid mollusc extract in rodent models for colorectal cancer", Molluscs 2009 Conference, Brisbane, Qld, Australia, November 25- 27, 2009, Poster

<u>Esmaeelian, B</u>, Benkendorff, K and Abbott, CA "Anti-cancer activity of the purified bioactive compounds from Australian muricid mollusk on colorectal cancer cell lines" The 13th International Symposium on Marine Natural Products, Phuket, Thailand, October 17-22, 2010, Poster & Oral presentation

<u>Esmaeelian, B</u>, Abbott, CA, Benkendorff, K, Le Leu, RK and Johnston, MR "*In vitro* and preclinical testing of purified muricid mollusc extract for colorectal cancer, Postgraduate Conference, School of Biological Sciences, Flinders University, Adelaide, Australia, June 2012, Oral presentation

<u>Esmaeelian, B</u>, Benkendorff, K, Le Leu, RK and Abbott, CA "The preclinical effects of purified mollusk extracts in a short term mouse model for early-stage colon cancer" Eleventh Annual AACR International Conference on Frontiers in Cancer Prevention Research, Anaheim, California, October 16-19, 2012, Poster

Acknowledgments

First of all, I would like to sincerely thank my supervisors Assoc. Prof. Catherine Abbott, for invaluable guidance, advice, help and support during my PhD journey, Dr. Kirsten Benkendorff, for impressive advise, continued support and guidance in most parts of my PhD and also great feedback on the draft thesis, Dr. Richard Le Leu for great advice in my *in vivo* experiments, and Assoc. Prof. Martin Johnston for precious help and advice on the chemical analysis of my compounds. You have all helped me on my PhD both on academic and personal level and for this I am truly grateful.

My special appreciation goes to School of Biological Sciences, Faculty of Science and Engineering, Flinders University for provision of the PhD scholarship, with top-up funding from a philanthropic grant to Dr Benkendorff.

I also would like to thank many people in Flinders University for their assistance and support. In the School of Biological Sciences, I am grateful to Dr. Chantel Westley, for her good advice and supplying the snails in the beginning of my study, David Rudd and Samsu Udayana Nurdin for their support and friendship, Dr. Hanna Krysinska and Ms Lisa Pogson for their help during my *in vivo* work and Assoc. Prof. Kathy Schuller for housing equipment and facilitating access to her lab.

In School of Medicine, my thank goes to Assoc. Prof. Peta Macardle from the flow cytometry analysis lab, Dr. Tim Chataway and Ms Nusha Chegeni from proteomics facility lab, Ms Roshini Somashekar, Joanne Wilkins and Jean Winter from gastrointestinal lab for their assistance and technical advice during my animal trial. I would also like to give special thanks to Dr. Daniel Jardine from the Flinders Analytical Laboratory, Faculty of Science and Engineering for his help in analysing my samples.

Finally, I would like to thank my parents and brother for their endless support and encouragement in these past 4 years.

Abbreviations

AARGC	acute apoptotic response to genotoxic carcinogens
ACF	aberrant crypt foci
ACS	American Cancer Society
AIF	apoptosis inducing factor
ALP	alkaline phosphatase
ALT	alanine aminotransferase
AOM	azoxymethane
ANOVA	analysis of variance
APC	adenomatous polyposis coli
AST	asparate aminotransferase
CDK	cyclin dependent kinase
CE	crude extract
CIMP	CpG island methylator phenotype
CIN	chromosomal instability
COX2	cyclo-oxygenase-2
CRC	colorectal cancer
DAPI	4',6-diamidino-2-phenylindole
DCM	dichloromethane
DMEM	Dulbecco's Modified Eagle's Medium
DMSO	dimethylsulphoxide
DNA	deoxyribonucleic acid
EC	European Commission
EGFR	epidermal growth factor receptor
EM	egg mass
EMEA	European Medicines Evaluation Agency
FDA	Food and Drug Administration
FOBT	fecal occult blood test
GC/MS	gas chromatography-mass spectrometry
GSK-3	glycogen synthase kinase-3
HCAs	heterocyclic amines
Hct	hemoglobin, hematocrit
HG	hypobranchial gland

HPLC	high-performance liquid chromatography
HX	hexane
IARC	International Agency for Research on Cancer
IBD	inflammatory bowel disease
LC/MS	liquid chromatography-mass spectrometry
LDH	lactate dehydrogenase
MCH	mean corpuscular hemoglobin
MCHC	mean corpuscular hemoglobin concentration
MCV	mean corpuscular volume
MMR	mismatch repair
MSI	microsatellite instability
MTT	3-(4, 5-dimethyl-thiazol-2-yl)-2,5-diphenyltetrazolium bromide
NMR	nuclear magnetic resonance
NCI	National Cancer Institute
NSAIDs	non-steroidal anti-inflammatory drugs
PAHs	polycyclic aromatic hydrocarbons
PBS	phosphate buffered saline
PI	propidium iodide
SEER	surveillance, epidemiology and end results
SRB	sulphorhodimine B reagent
Str	staurosporine
TLC	thin-layer chromatography
TNF	tumour necrosis factor
TNM	tumor node metastasis
TUNEL	terminal deoxynucleotidyl transferase dUTP nick end labeling
TYR	tyrindoleninone
VEGF	vascular endothelial growth fact