Pre-Clinical Multi-Modal Imaging for Assessment of Pulmonary Structure, Function and Pathology





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Pre-Clinical Multi-Modal Imaging for Assessment of Pulmonary Structure, Function and Pathology

by

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Abstract

In this thesis, we describe several imaging techniques specifically designed and developed for the assessment of pulmonary structure, function and pathology. We then describe the application of this technology within appropriate biological systems, including the identification, tracking and assessment of lung tumors in a mouse model of lung cancer.

The design and development of a Large Image Microscope Array (LIMA), an integrated whole organ serial sectioning and imaging system, is described with emphasis on whole lung tissue. This system provides a means for acquiring 3D pathology of fixed whole lung specimens with no infiltrative embedment medium using a purpose-built vibratome and imaging system. This system enables spatial correspondence between histology and non-invasive imaging modalities such as Computed Tomography (CT), Magnetic Resonance Imaging (MRI) and Positron Emission Tomography (PET), providing precise correlation of the underlying "ground truth" pathology back to the *in vivo* imaging data. The LIMA system is evaluated using fixed lung specimens from sheep and mice, resulting in large, high-quality pathology datasets that are accurately registered to their respective CT and H&E histology.

The implementation of an *in vivo* micro-CT imaging system in the context of pulmonary imaging is described. Several techniques are initially developed to reduce artifacts commonly associated with commercial micro-CT systems, including geometric gantry calibration, ring artifact reduction and beam hardening correction. A computer controlled Intermittent Iso-pressure Breath Hold (IIBH) ventilation system is then developed for reduction of respiratory motion artifacts in live, breathing mice. A study validating the repeatability of extracting valuable pulmonary metrics using this technique against standard respiratory gating techniques is then presented.

The development of an *ex vivo* laser scanning confocal microscopy (LSCM) and an *in vivo* catheter based confocal microscopy (CBCM) pulmonary imaging technique is described. Direct high-resolution imaging of sub-pleural alveoli is presented and

an alveolar mechanic study is undertaken. Through direct quantitative assessment of alveoli during inflation and deflation, recruitment and de-recruitment of alveoli is quantitatively measured. Based on the empirical data obtained in this study, a new theory on alveolar mechanics is proposed.

Finally, a longitudinal mouse lung cancer study utilizing the imaging techniques described and developed throughout this thesis is presented. Lung tumors are identified, tracked and analyzed over a 6-month period using a combination of micro-CT, micro-PET, micro-MRI, LSCM, CBCM, LIMA and H&E histology imaging. The growth rate of individual tumors is measured using the micro-CT data and traced back to the histology using the LIMA system. A significant difference in tumor growth rates within mice is observed, including slow growing, regressive, disappearing and aggressive tumors, while no difference between the phenotype of tumors was found from the H&E histology. Micro-PET and micro-MRI imaging was conducted at the 6-month time point and revealed the limitation of these systems for detection of small lesions (<2mm) in this mouse model of lung cancer. The CBCM imaging provided the first high-resolution live pathology of this mouse model of lung cancer and revealed distinct differences between normal, suspicious and tumor regions. In addition, a difference was found between control A/J mice parenchyma and Urethane A/J mice 'normal' parenchyma, suggesting a "field effect" as a result of the Urethane administration and/or tumor burden. In conclusion, a comprehensive murine lung cancer imaging study was undertaken, and new information regarding the progression of tumors over time has been revealed.

Declaration

I certify that this thesis does not incorporate without acknowledgment any material previously submitted for a degree or diploma in any university; and that to the best of my knowledge and belief it does not contain any material previously published or written by another person except where due reference is made in the text.

NY

Eman Namati

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List of Symbols and Abbreviations

<i>v</i>	Frequency
λ	Wavelength
18F-FDG	Fluedeoxyglucose F18
18F-FLT	Fluorothymidine F18
A/D	Analog-to-Digital
AOTF	Acousto-Optical Tunable Filter
BASC	Bronchio-Alveolar Stem Cell
<i>c</i>	Speed
CBCM	Catheter Based Confocal Microscopy
CCD	Charge Coupled Device
cGy	Centi-Gray
СТ	Computed Tomography
DPSS	Double Pumped Solid State
dsDNA	Double Stranded DNA
<i>E</i>	Energy
EM	Electro Magnetic
eV	electron Volt
FFT	Fast Fourier Transform
FOV	
GFP	Green Fluorescent Protein
GRE	Gradient-Recalled Echo
GRIN	Gradient-Index
HU	Hounsfield Unit
IFFT	Inverse Fast Fourier Transform
IIBH	Intermittent Iso-pressure Breath hold
IM	Intra Muscular
IP	Intra Peritoneal
IV	Intra Venous
LE	Late Expiratory
LI	Late Inspiratory
LIMA	Large Image Microscope Array
LSCM	Laser Scanning Confocal Microscopy

MCL	Mean Chord Length
MCLa	
MCLw	Mean Chord Length of Wall
MEMS	Micro Electro Mechanical System
micro-CT	micro-Computed Tomography
MRI	
MTF	
NSCLC	Non-Small-Cell Lung Cancer
PBS	Phosphate Buffer Saline
PID	Proportional Integrative Derivative
РМТ	Photo Multiplier Tube
RECIST	Response Evaluation Criteria in Solid Tumors
rtTA	
SCLC	Small-Cell Lung Cancer
SNR	Signal to Noise Ratio
ssDNA	
TTL	Transistor-Transistor Logic
WHO	World Health Organization